

Microalgal taxonomic diversity from shallow acidophilus freshwater bodies from the south of Chile

Diversidad taxonómica de microalgas en cuerpos dulceacuícolas someros y acidófilos del sur de Chile

Mariela A. González^{1*}, Ingrid de L. Inostroza¹, Visitación Conforti² & Enrique Ascencio¹

¹Laboratorio de Cultivo de Microalgas (FICOLAB), Departamento de Botánica, Facultad de Ciencias Naturales y Oceanográficas, Universidad de Concepción, Chile.

²Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales, Instituto de Biodiversidad y Biología Experimental y Aplicada, CONICET-Universidad de Buenos Aires, Buenos Aires, Argentina.

*Email: mgonzale@udec.cl

ABSTRACT

This research is the first taxonomic approach about the microalgae inhabiting shallow acidophilus freshwater bodies (Chapito pool, "small dam" and Tepual stream) within the Katalapi Park, X Region, Chile. A total of 80 taxa were identified based on morphocytological attributes, being the Streptophyta-Zygnematophyceae the most abundant group with a dominance of Desmidiaceae (47 species), follow by Cyanobacteria (14), Euglenophyceae (12), Chlorophyta-Chlorophyceae (5), and Dinophyceae and Ochrophyta-Raphidophyceae with one taxon each group. The genera with the higher number of species were *Cosmarium* (12), *Closterium* (11) and *Staurastrum* (8). The pH ranged from 5.0 to 6.1 during the recollection period. Most of the desmids species identified (70.2 %) are from acidophilus environments, 48.6 % are indicators of mesotrophy and 21.6% of meso-oligotrophic habitats. Twenty-four species are cited for the first time for Chile. Although the study was restricted to a relatively small area, the results suggest that a great diversity of microalgae might be also found in similar ecological areas within the country.

Keywords: blue-greens, desmids, morphology, photosynthetic euglenoids, taxonomy.

RESUMEN

Esta investigación es el primer aporte taxonómico sobre las microalgas que habitan en cuerpos de agua acidófilos, someros (Laguna Chapito, "pequeño tranque" y estero Tepual) al interior del Parque Katalapi, X Región, Chile. Se identificó un total de 80 taxa basado en atributos morfológicos, siendo el grupo Streptophyta-Zygnematophyceae el más abundante con dominancia de Desmidiaceae (47 especies), seguido por Cyanobacteria (14), Euglenophyceae (12), Chlorophyta-Chlorophyceae (5), y Dinophyceae y Ochrophyta-Raphidophyceae con un taxón cada grupo. Los géneros con el mayor número de especies fueron *Cosmarium* (12), *Closterium* (11) y *Staurastrum* (8). El rango de pH durante el periodo de recolección fluctuó entre 5,0 a 6,1. La mayoría de las especies de desmidiaceas identificadas (70,2 %) son de ambientes acidófilos, el 48,6 % son indicadoras de mesotrofia y el 21,6% de hábitats meso-oligotróficos. Veinticuatro especies se citan por primera vez para Chile. Aunque el estudio estuvo restringido a un área relativamente pequeña, los resultados sugieren la potencialidad de encontrar una gran diversidad de microalgas en áreas ecológicas similares en el país.

Palabras clave: algas verde-azules, Desmidiaceas, euglenoides fotosintéticos, morfología, taxonomía.

INTRODUCTION

The taxonomic knowledge about the microalgae inhabiting water bodies (pools, ponds, streams, rivers) within national parks in the last 50 years in Chile, is scarce. Several studies have been done in the National Park "Torres del Paine" (Soto *et al.* 1994, Campos *et al.* 1994a, b, Campos *et al.* 1996, De los Ríos & Soto 2009) and in the northern Patagonian lakes Chapo (Villalobos *et al.* 2003) and Riñihue (Woelfl *et al.* 2003): however, most of them from a limnological point of view. So, only chlorophyll "a" or a list of the main phytoplankton species found in these water bodies were included. More recently, Palau & Caputo (2013), characterized limnologically 15 pristine water bodies from the "Fundación San Ignacio de Huinay" area in the north Patagonia (X Region), where they identified 60 taxa of microalgae but mostly (70%) at generic level. The only studies focused on the biodiversity of microalgae were those from Parra & González (1977, 1978) on some lakes and other freshwater bodies (i.e. ponds, streams) from Chiloe Island, Theoduloz & Parra (1984) on the Lake Systems of Cordillera del Paine, and Williamson (2004) from Laguna San Rafael National Park. The diatoms have been studied by Rose & Cox (2002) from different benthic habitats within seven upland lakes in the Laguna San Rafael National Park and the presence of specific species by Rivera *et al.* (2002), in Lake Chungara (*Stephanodiscus agassizensis* Hakansson & Kling), Rivera & Cruces (2008) in Lauca River (both from Parque Nacional Lauca) (*Fragilaria capucina* var. *mesogongyla* Frenguelli) and by the same later authors (2015) in Salar Aguas Calientes and Salar de Huasco (Parque Nacional Salar del Huasco) (*Frankophila sudamericana* Rivera & Cruces).

The present study was carried out in acidophilus freshwater bodies (a pond, a small reservoir and a stream) within the Katalapi Park, a protected area for more than two decades, with the aim to know the most common microalgae (excl. Diatoms) inhabiting those environments.

MATERIAL AND METHODS

The Katalapi Park (41° 31'12"S; 72° 45'42"W) is located within the Ecoregion of the Valdivian Temperate Rain Forest, which has a high degree of endemism. It is a private protected area since 1994, created to support scientific research linked to the native forest. The park has 28 ha (about 56 acres) of native forest and open fields. Water bodies are abundant in the park (streams, a pool and ponds).

The water bodies under study were: the Chapito pool, a very shallow pool of approximately 20 to 80 cm depth and 30 m length x 10 m wide (Fig. 1a); a "small dam" of 10 m length x 5 m wide with a depth fluctuating between 30 cm to 2 m (Fig. 1b); and the Tepual stream (Fig. 1c). Phytoplankton from these three water bodies were collected with the help of a plankton net (45 µm mesh diameter) during an annual period, including the four seasons of the year (October 2016 and February, May and August 2017). Water temperature (current thermometer) and pH (pH paper) data were also taken.

The samples were observed and photographed *in vivo* with the help of a standard (Olympus CX31) and inverted light (Olympus CKX 41) microscope with a camera (518CU 5.0M CMOS) attached to them, and using an *ad hoc* software to capture images and obtain micron size standards. Later on the samples were preserved in formalin 4% and stored at FICOLAB (Botany Department, University of Concepcion). The species were identified based on the traditional morphological (i.e. shape, size) and cytological observations (i.e. shape, color and number of chloroplasts, presence/absence of pyrenoid, number and location of nucleus, when visible). The following identification keys were used: Cyanobacteria are given according to Komárek & Anagnostidis (2000, 2005), Komárek & Komárková (2002), with some modification based on recent data for the genus *Planktothrix* Gaget *et al.* (2015) and taxonomic changes at the generic level in *Geitlerinema*, Strunecký *et al.* (2017); Euglenophyceae are presented according to Ciugulea & Triemer (2010), Marin *et al.* (2003), Tell & Conforti (1986) and Hübber Pestalozzi (1955); Ochrophyta-Raphidophyceae according to Wehr & Sheath (2003); Dinophyceae based on Carty (2014); Chlorophyta-Chlorophyceae, according to Nakasawa *et al.* (2004) for the genus *Asterococcus*, Tsarenko (2011) for the genus *Fusola* and Pröschold (pers. comm. 2018) for the genus *Chlamydocapsa*, and finally, Streptophyta-Zygnematophyceae based on Sampaio (1944), Hirano (1955-1960), Grönblad *et al.* (1968), Prescott *et al.* (1981), Wehr & Sheath (2003) and John *et al.* (2008).

RESULTS

The pH values ranged between 5.0-5.8 in Chapito pool, 5.8-6.0 in the "small dam" and 5.5 - 5.9 in the Tepual stream, during the annual recollection period. The temperatures, on the other hand, fluctuated from 7.2 to 18 °C in Chapito pool, 7 to 14.8 °C in the "small dam" and 6.5 to 12.5°C in the Tepual stream.

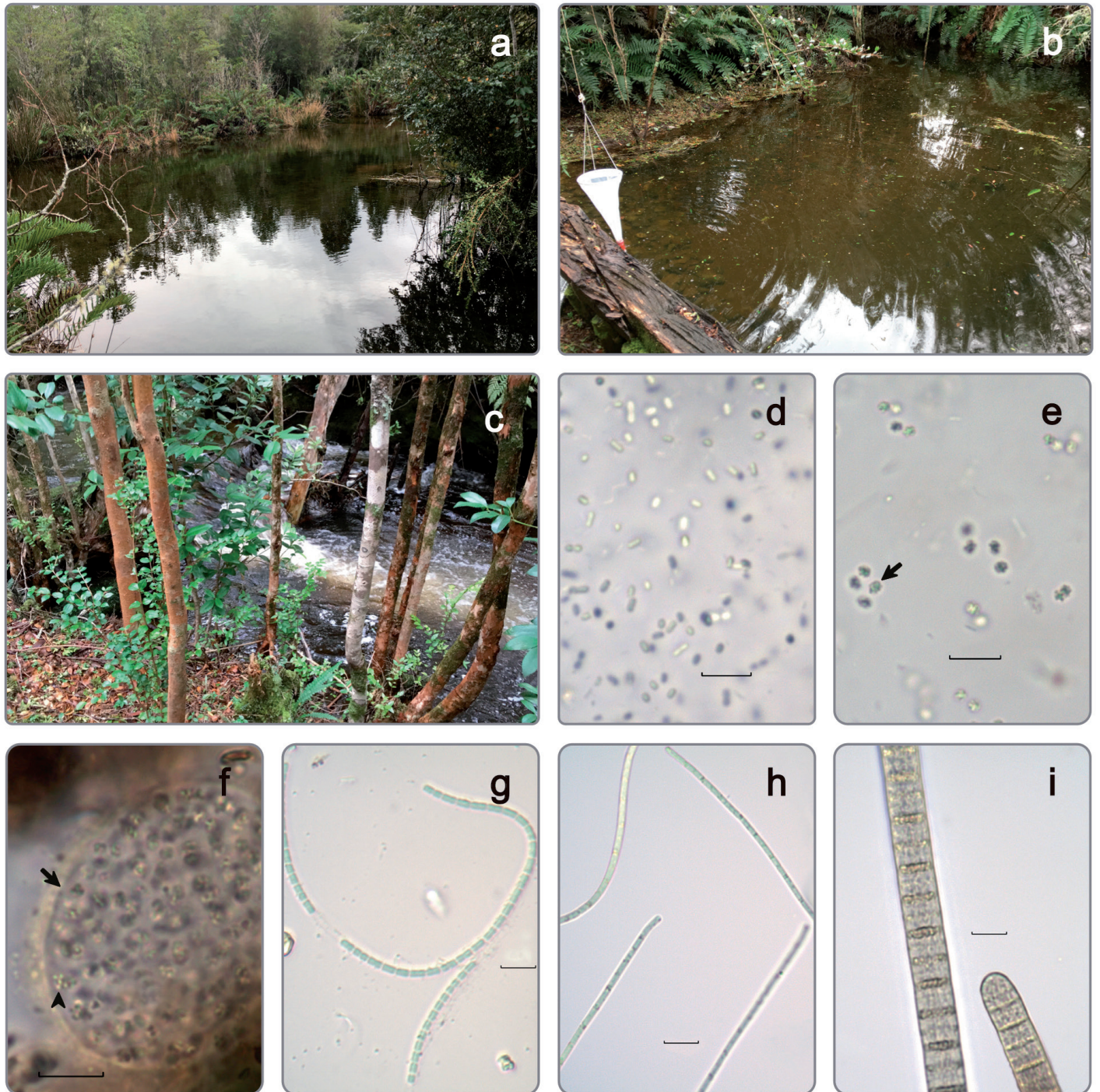


FIGURE. 1 (a-c) Places of collection. (a) Chapito pool, (b) small dam, (c) Tepual stream; (d) *Anathece minutissima*, detail of the cenobium; (e) *Microcystis natans*, detail of the cenobium, note aerotopes (arrow); (f) *Microcystis firma*, note firm mucilaginous envelop (arrow) and aerotopes (arrowhead); (g) *Pseudanabaena minima*; (h) *Anagnostidinema cf. amphibium*; (i) *Tychonema cf. bornetii*. Scale bar: 10 μm ./ (a-c) Lugares de colecta. (a) Laguna Chapito, (b) Repesita, (c) Riachuelo Tepual; (d) *Anathece minutissima*, detalle del cenobio; (e) *Microcystis natans*, detalle del cenobio, note presencia de aerótopos (flecha); (f) *Microcystis firma*, note firme envoltura mucilaginosa (flecha) y aerótopos (punta flecha); (g) *Pseudanabaena minima*; (h) *Anagnostidinema cf. amphibium*; (i) *Tychonema cf. bornetii*. Barra escala: 10 μm .

A total of 80 taxa were identified, being the Streptophyta-Zygnematophyceae the most abundant group with a dominance of Desmidiaceae (47 species). The genera with the higher number of species were *Cosmarium* (12), *Closterium* (11) and *Staurastrum* (8). The presence of the other taxonomic groups was as followed: Cyanobacteria with 14 taxa, Euglenophyceae with 12 taxa, Chlorophyta-Chlorophyceae with 5 taxa, and Dinophyceae and Ochrophyta-Raphidophyceae with one taxon each group.

The taxonomic identification is as followed:

Cyanobacteria

Order Chroococcales

Family Synechococcaceae

Anathece minutissima (West) Komárek, Kastovsky & Jezberová 2011

Coenobia microscopic, gelatinous, elongate (125 µm x 62.5 µm), enclosed by a fine, colorless, homogeneous mucilage with irregularly and more or less densely aggregated cells. Cells broadly oval or rod-shaped (0.8-1.0 µm x 1.5 µm), greyish in color and with homogeneous content (Fig.1d).

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Family Chroococcaceae

Chroococcus minutus (Kuetzing) Naegeli 1849

Coenobia with 4-8 cells enclosed by a homogeneous, hyaline sheath. Cells spherical or hemispherical after division, 4-6 µm in diameter, blue-greens, with granulate content.

Recorded for Chile (see Parra *et al.* 1982a).

Habitat: Chapito pool (August 2017).

Limnococcus limneticus (Lemmermann) Komárková, Jezberová, O. Komárek & Zapomelová 2010

Coenobia with 4-8 cells irregularly distributed or in groups of 2 to 4, enclosed by a wide, colorless, hyaline gelatinous envelope. Cells spherical, 5 µm in diameter.

Recorded for Chile as *Chroococcus limneticus* Lemm. (see Parra *et al.* 1982a).

Habitat: Chapito pool (August 2017).

Family Merismopediaceae

Aphanocapsa elachista W. & G.S. West 1894

Coenobia 80 - 110 µm in diameter, with cells sparsely disposed, forming pairs after division. Cells spherical, with homogeneous content, 1.5-2.0 µm in diameter.

Recorded for Chile (see Almanza *et al.* 2016; as *Microcystis elachista* (W. et G.S. West) Starmach, see Parra *et al.* 1982a).

Habitat: Chapito pool (February 2017).

Aphanocapsa incerta (Lemmermann) Cronberg et Komárek 1994

Coenobia 50-60 µm in diameter, with a great number of cells densely aggregated. Cells spherical, 1.8-2.8 µm in diameter, with homogeneous content.

Recorded for Chile (see Almanza *et al.* 2016; as *Microcystis incerta* Lemm., see Parra *et al.* 1982a).

Habitat: Chapito pool (February 2017).

Family Microcystaceae

Microcystis natans Lemmermann ex Skuja 1934

Coenobia microscopic, spherical to elongate, 60-250 µm in diameter, with cells irregularly, not very densely packed. Mucilage colorless, homogeneous, fine. Cells spherical (1.5 µm) to oval before division (2 µm), with aerotopes (Fig.1e).

Recorded for the first time for Chile.

Habitat: Chapito pool (February 2017).

Microcystis firma (Kuetzing) Schmidle 1902

Coenobia microscopic, spherical or sub-spherical, 40-50 µm in diameter, with densely packed cells, enclosed by a mucilaginous envelop forming a distinct margin. Cells 1.7-2.1 µm in diameter, with aerotopes (Fig.1f).

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Family Pseudanabaenaceae

Pseudanabaena catenata Lauterborn 1915

Trichomes 1.5 µm wide; cells cylindrical, 4-5 µm long.

Recorded for Chile (see Almanza *et al.* 2016; Parra *et al.* 1982a).

Habitat: Chapito pool (February 2017).

Pseudanabaena minima (G.S. An) Anagnostidis 2001

Trichomes shorts, straights or curves, pale blue-greens, 1.8-2 µm wide. Cells isodiametric or longer than wide (2-4 µm long). Cellular content homogeneous (Fig.1g).

Recorded for the first time for Chile.

Habitat: Chapito pool and "small dam" (February 2017).

Order Oscillatoriales

Family Coleofasciculaceae

Anagnostidinema cf. ***amphibium*** (Agardh ex Gomont) Strunecky, Bohunická, Johansen et Komárek 2017.

Trichomes thins, longs, motile, light blue-greens, usually straight or slightly curves, not constricted at the cross-walls, 1.5-1.8 µm wide, not or very slightly attenuated at the ends, not capitated. Cells 4-8 µm long, pale blue-greens, with commonly one notorious refractive granule at either side of the cross-walls. Apical cell rounded, without calyptra.

Necridial cells were not observed (Fig.1h).

Initially, we identified this taxon as *Geitlerinema* cf. *unigranulatum* (R. N. Singh) Komárek et Azevedo 2000. However, Bittencourt-Oliveira *et al.* (2009) suggested, based on morphology, ultrastructure and DNA data [sequence of the *pcB-cpcA* intergenic spacer (PC-IGS)] that *G. unigranulatum* is synonymous of *Geitlerinema amphibium* (C. Agardh ex Gomont) Anagnostidis 1989. Later on, Strunecký *et al.* (2017) revised the genus *Geitlerinema* and created a new genus, *Anagnostidinema* and included *G. amphibium* as synonymous of *Anagnostidinema amphibium*. To validate the identity of this taxon, molecular data is needed.

Recorded for the first time for Chile.

Habitat: Chapito pool (October 2016).

Family Oscillatoriaceae

Oscillatoria curviceps Agardh ex Gomont 1892

Trichomes blue-green, 10 µm in diameter, slightly curved at the extremes which can or cannot be moderately attenuated, not constricted at the cross walls, where granules are usually present. Cells very short, 1.8-3.5 µm. Apical cells widely round, not capitated, sometimes with thickening of the outer wall.

Recorded for Chile (see Parra *et al.* 1982a).

Habitat: Chapito pool (October 2016, February, May and August 2017).

Family Phormidiaceae

Tychonema cf. *bornetii* (Zucal) Anagnostidis et Komárek 1988

Trichomes brownish-green, straight, cylindrical, isopolar, not attenuated to the ends, 11-12 µm width, not constricted at the cross walls. Cells more or less isodiametric or slightly longer or shorter than wide (10-12 µm in length), notoriously granulated at the cross walls, and less notorious and more disperse granulation within the protoplasm. Apical cells broadly rounded with slightly thickened outer cell wall (Fig.1i). Even though most of the morphological attributes observed correspond with those of *T. bornetii*, molecular studies are needed to confirm this identification.

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Planktothrix cf. *clathrata* (Skuja) Anagnostidis & Komárek 1988

Trichomes blue-green, cylindrical, not narrowed to the ends, 6 to 9 µm in diameter, with distinct constrictions at the cross walls. Cells always shorter than longer (2 to 4 µm in length), with aerotopes. Apical cell rounded, not capitated (Fig. 2a).

Even though most of the morphological attributes observed correspond with those of *P. clathrata*, molecular studies are needed to confirm this identification.

Recorded for the first time for Chile.

Habitat: Chapito pool (February 2017).

Planktothrix cf. *suspensa* (Pringsheim) Anagnostidis et Komárek 1988

Trichomes blue-green, cylindrical, straight, 4-5 µm in width, not constricted at cross walls. Cells usually ± isodiametric, or shorter and sometimes longer than wide (4-7 µm in length), with aerotopes. Apical cell rounded, in some individuals with slight thickness of the outer wall. As in the case of *Planktothrix* cf. *clathrata*, molecular studies are needed to confirm the identification of this taxon (Fig. 2b).

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Dinoflagellata

Order Peridiniales

Parvodinium sp.

Cells small, 22-29 µm long, 19-22 µm wide, ovoid, with numerous plastids yellowish-brown in color. The epitheca is bigger than the hypotheca; cingulum, wide. Apical pore visible. Plates thin, not visible in living organisms (Fig. 2c). This taxon probably corresponds to *P. cf. umbonatum* (Stein) Carty 2008 (E. Ascencio, pers.observ.).

The genus has been recorded for Chile as *Peridinium* [*Peridium inconspicuum* Lemm., Thomasson 1963, now *Parvodinium inconspicuum* (Lemm.) Carty].

Habitat: Chapito pool (February and August 2017).

Ochrophyta - Rhaphidophyceae

Order Chattonellales

Vacuolaria virescens Cienkowski 1870

Cells biflagellated, ovoid to ellipsoid in frontal view, 52-54 µm long, 38-46 µm wide. Chloroplasts numerous, discoid, light-green in color. Nucleus ellipsoidal (app. 10 µm size) located in the center of the cell. Many refractile granules present in the cell periphery (Fig 2d).

Recorded for the first time for Chile.

Habitat: "small dam" (February 2017).

Euglenophyta - Euglenophyceae

Order Euglenales

Euglena intermedia (Klebs) Schmitz 1884 (= *Euglena deses* var. *intermedia* Klebs, 1883)

Cells metabolic, cylindrical with the anterior end blunt and the posterior tapering in a rounded tail, 105-120 µm long, 10-15 µm wide. Pellicle finely striated. Chloroplasts numerous, discoid without pyrenoid; paramylon bodies rod-like. Flagellum about one sixth of the body length. Eyespot conspicuous.

Recorded for Chile (see Parra *et al.* 1982b).

Habitat: Chapito pool (Febrero 2017).

Euglena mutabilis Schmitz 1884

Cells metabolic, elongate, slender, almost cylindrical, with the anterior end rounded and the posterior sharpened, 64-70 μm long, 5-9 μm wide. Pellicle spirally striated; numerous plate-like chloroplasts, curved, with naked pyrenoid; small paramylon grains and central nucleus.

Recorded for Chile (see Parra *et al.* 1982b).

Habitat: Chapito pool (August 2017).

Euglena oblonga Schmitz 1884

Cells metabolic, spindle shaped, with anterior end rounded and the posterior ending in a blunt point or extended into a short tail, 41-50 μm long, 21-27 μm wide. Pellicle spirally striated. Chloroplasts numerous, deeply incised with long ribbon-shaped lobes and diplopyrenoids (Fig. 2e).

Recorded for the first time for Chile.

Habitat: Chapito pool (February and August 2017).

Lepocinclis spirogyroides Marin & Melkonian 2003

Cells cylindrical or flattened with the anterior end rounded and the posterior ending in a colorless pointed tail, 75-90 μm long, 9-14 μm wide. Yellow-brown pellicle spirally striated, strips with brick-like ornamentation. Numerous small, disc-shaped chloroplasts. Two large paramylon bodies one anterior and one posterior to the nucleus, and smaller ones disperse within the cell (Fig. 2f).

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Lepocinclis steinii Lemmermann 1901

Cells fusiform with the anterior end truncate and the posterior with short conical caudal appendix, 27-30 μm long, 12-16 μm wide. Pellicle longitudinally striated. Chloroplasts numerous, discoid. Two large lateral paramylon bodies (Fig. 2g).

Recorded for the first time for Chile.

Habitat: "small dam" (October 2016).

Monomorphina pyrum (Ehrenberg) Mereschkowsky *emend.* Kosmala et Zakrys 2007

Pear shaped cells 30-34 μm long, 15-21 μm wide; anterior end symmetric and posterior end with a long straight and finely pointed tail, 10-12 μm long. Pellicle with laevogyrate spirally keels. One orbicular perforated chloroplast. Two lateral paramylon plates.

Recorded as *Phacus pyrum* (Ehr.) Stein (see Dürschmidt 1978).

Habitat: "small dam" (October 2016).

Trachelomonas abrupta Swirenko *emend.* Deflandre 1926

Lorica 30-34 μm long, 15-21 μm wide, ellipsoidal, with sides more or less curved, sometimes with a quadrangular tendency in longitudinal optical view, yellowish-brown, punctate, covered with minute spines, difficult to see.

Recorded for Chile (see Parra & González 1978).

Habitat: Chapito pool (February 2017).

Trachelomonas hispida (Perty) Stein *emend.* Deflandre 1926

Lorica 23-33 μm long 19-24 μm wide, broadly ellipsoidal, reddish-brown, punctate, covered with short conical spines, irregularly distributed; apical pore rounded by an annular thickening.

Recorded for Chile (see Pereira 2000; Parra *et al.* 1982b).

Habitat: Chapito pool (February and August 2017).

Trachelomonas intermedia Dangeard 1901

Lorica 9-22 μm long, 14-20 μm wide, sub-spherical or ellipsoidal, yellowish-brown, finely punctate; apical pore rounded by an annular thickening.

Recorded for Chile (see Dürschmidt 1978).

Habitat: Chapito pool (August 2017).

Trachelomonas pulcherrima Playfair 1915

Lorica 20-24 μm long, 9-12 μm wide, ellipsoidal, very elongated, with poles broadly rounded, the posterior sometimes acuminate, light-yellow to brown, finely punctate.

Recorded for Chile (see Parra & González 1978).

Habitat: Chapito pool (August 2017).

Trachelomonas raciborskii Woloszyńska 1912

Lorica 30-40 μm long, 24-30 μm wide, ellipsoidal, thick, dark-brown, punctate, with conical and strong spines only close to the poles. Apical pore without a collar and surrounded by spines (Fig. 2h).

Recorded for the first time for Chile.

Habitat: Chapito pool (February 2017).

Trachelomonas rotunda Svirenko *emend.* Deflandre 1926

Lorica 20-30 μm long, 19-30 μm wide, sub-spherical or broadly ellipsoidal, thick, dark-brown, strongly punctate; sometimes, with very short spines in its anterior region. Apical pore surrounded by an annular thickening (Fig. 2i).

Recorded for the first time for Chile.

Habitat: Chapito pool (Agosto 2017).

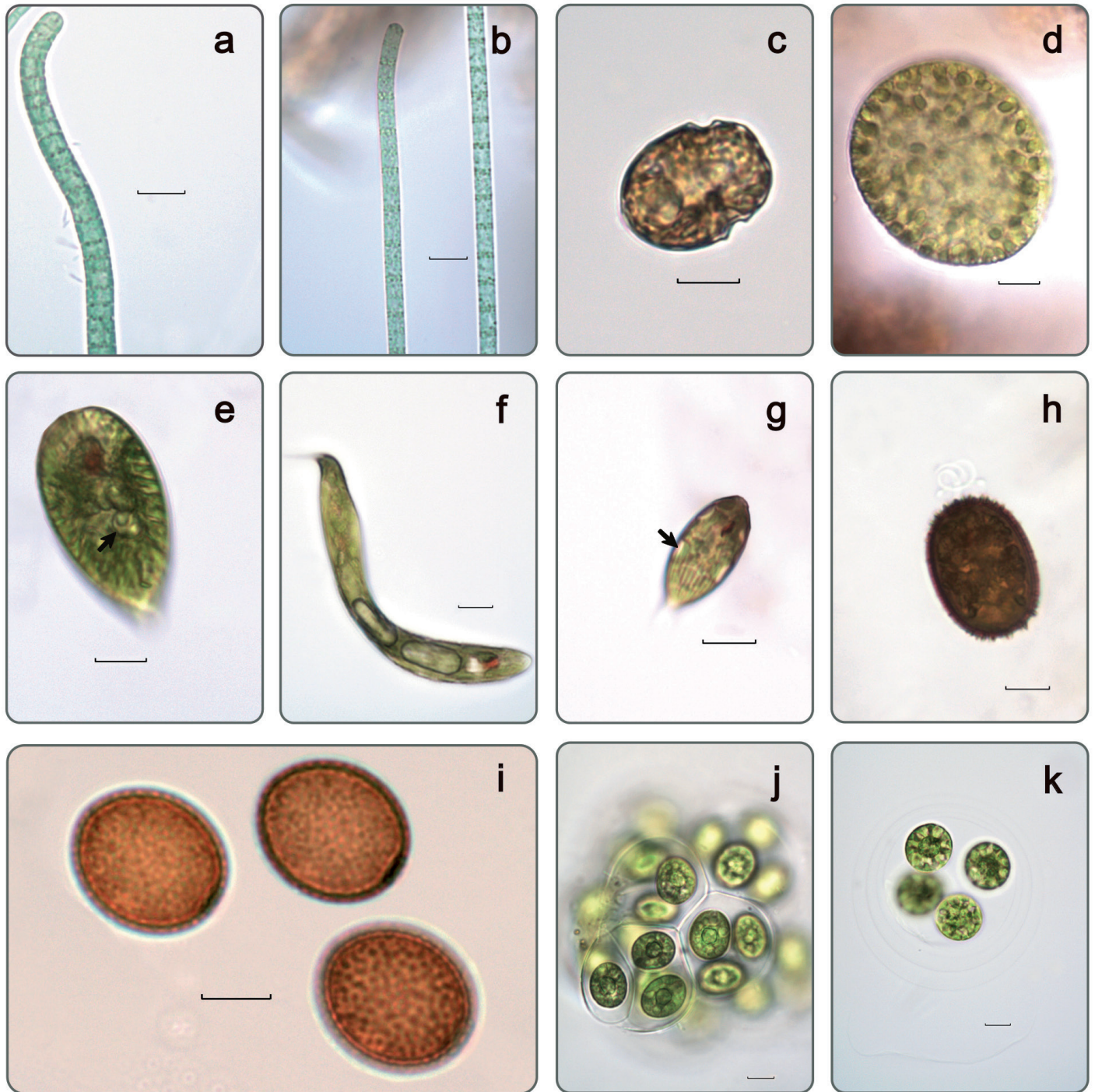


FIGURE 2. (a) *Planktothrix* cf. *clathrata*; (b) *Planktothrix* cf. *suspensa*; (c) *Parvodinium* sp.; (d) *Vacuolaria virescens*; (e) *Euglena oblonga*, diplopyrenoid (arrow); (f) *Lepocinclis spirogyroides*; (g) *Lepocinclis steinii*, paramylon (arrow); (h) *Trachelomonas raciborskii*; (i) *Trachelomonas rotunda*; (j) *Asterococcus* cf. *limneticus*; (k) *Asterococcus* cf. *superbus*. Scale bar: 10 µm. / (a) *Planktothrix* cf. *clathrata*; (b) *Planktothrix* cf. *suspensa*; (c) *Parvodinium* sp.; (d) *Vacuolaria virescens*; (e) *Euglena oblonga*, diplopyrenoid (flecha); (f) *Lepocinclis spirogyroides*; (g) *Lepocinclis steinii*, paramylon (flecha); (h) *Trachelomonas raciborskii*; (i) *Trachelomonas rotunda*; (j) *Asterococcus* cf. *limneticus*; (k) *Asterococcus* cf. *superbus*. Barra escala: 10 µm.

Chlorophyta – Chlorophyceae

Orden Chlamydomonadales

Asterococcus cf. *limneticus* G. M. Smith 1918

Coenobia 45 to 100 µm in diameter, composed of spherical to ellipsoidal cells (16-20 µm long x 12-15 µm wide), forming groups of 4 to 16 cells embedded in a mucilage. Chloroplast, one, stellate, with a central pyrenoid (Fig. 2j).

According to Nakazawa *et al.* (2004) and based on morphological attributes this taxon belongs to *Asterococcus limneticus*; however, molecular data is needed to confirm its identity.

Recorded for the first time for Chile.

Habitat: Chapito pool (February, May 2017).

Asterococcus cf. *superbus* (Cienkowski) Scherffel 1908

Cells spherical, single or in small groups of four (17-30 µm in diameter) with a wide mucilaginous lamellate (4) envelope. Chloroplast one, central, with numerous arms radiating from a central pyrenoid (Fig. 2k).

As in the previous taxon, molecular data are needed to confirm the identity of this organism as *Asterococcus superbus*.

Recorded for Chile (see Parra *et al.* 1983a).

Habitat: Chapito pool (May 2017).

Chlamydocapsa cf. *lobata* Broady 1977

Cells spherical or sub-spherical, 8-10 µm in diameter, distributes in groups of 8 within a wide mucilaginous envelope. Presence of a fine mucilage around each cell or groups of cells. Chloroplast parietal, lobate with a central, basal pyrenoid (Fig. 3a).

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Chlamydocapsa cf. *planctonica* (West & G.S. West) Fott

Coenobia 60 to 85 µm in diameter, composed of numerous cells surrounded by a hyaline mucilaginous envelope. Cells irregularly distributed, ovoid, 10-14 (-17) µm long, 7-9 (-12) µm wide. Chloroplast parietal, cup-shaped, with one pyrenoid, spherical, sub-median in position (Fig. 3b).

Recorded for Chile (see Parra *et al.* 1983a).

Habitat: Chapito pool (May 2017), "small dam" (February 2017).

Order: Sphaeropleales

Fusola cf. *viridis* J. W. Snow 1903

Cells in groups of four alternately disposed within a colorless mucilage, fusiform, 30-35 µm long, 7-8 µm wide. Chloroplast parietal, laminar, with distinct central pyrenoid (Fig. 3c).

Recorded for the first time for Chile.

Habitat: Chapito pool (May 2017).

Streptophyta - Zygnematophyceae

Order Desmidiiales

Actinotaenium globosum (Bulnheim) Förster ex Compère 1976

Cells 31.3-38.3 µm long, 23.1-24.4 µm wide. Isthmus little marked 21.8-23.1 µm wide. Semicells globular, apex rounded. Cell wall dotted.

Recorded for Chile (see Williamson 2004) and as *Cosmarium globosum* Bulnheim (see Parra *et al.* 1983b).

Habitat: Chapito pool (August 2017), "small dam" (February 2017).

Bambusina brebisonii Kützing ex Kützing 1849

Cells barrel-shaped 25.1-27.8 µm long, 17-19.1 µm wide. Cells are united by their apices to form filaments; semicells pyramidal-truncated.

Recorded for Chile (see Williamson 2004; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, February and May 2017).

Closterium angustatum Kützing ex Ralfs 1848

Cells long, slightly curved, in the mid-region with parallel margins, toward the ends only slightly narrowed, 183-450 µm long, 13-23 µm wide. Apices are capitated. Chloroplasts with 3 to 5 longitudinal ridges; pyrenoids aligned. Cell wall with transverse bands and longitudinal ribs widely separated (Fig. 3d).

Recorded for the first time for Chile.

Habitat: Chapito pool (October 2016, February, May and August 2017), "small dam" (October 2016, February 2017).

Closterium ehrenbergii Meneghini ex Ralfs 1848

Cells large, moderately curved 400 µm long, 60-70 µm wide, swollen at the center and attenuating towards the rounded apices. Chloroplasts with 4-7 longitudinal ridges, with many pyrenoids scattered. Cell wall finally striated.

Recorded for Chile (see Pereira *et al.* 2000; Parra *et al.* 1983b).

Habitat: Chapito pool (February and August, 2017).

Closterium gracile Brébisson ex Ralfs 1848

Cells long, narrow, 140-190 µm long, 5 µm wide, slightly curved but straight at the center, tapered at both ends which become slightly incurved. Chloroplast with 4-7 pyrenoids aligned. Cell wall smooth.

Recorded for Chile (see Almanza *et al.* 2016; Williamson 2004; EULA 2013; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, February, May and August 2017), "small dam" (February 2017).

Closterium jenneri Ralfs 1848

Cells small, evenly curved, 65-80 µm long, 10 µm wide,

margins almost parallel in the central part that narrow slightly at the ends; apices rounded. Chloroplast with 3-5 longitudinal ridges, 2-5 pyrenoids in each semicell. Cell wall striated although sometimes difficult to detect (Fig. 3e).

Recorded for Chile (Borge 1906)

Habitat: Chapito pool (October 2016, February and August 2017).

Closterium kuetzingii Brébisson 1856

Cells 300 µm long, 13-20 µm wide, with a broad fusiform-lanceolate middle region, attenuating abruptly into long, narrow ends. Chloroplasts with 4-6 longitudinal ridges and 4-10 pyrenoids scattered. Cell wall finely striated.

Recorded for Chile (see Parra *et al.* 1983b, 1999, 2005; Williamson 2004; Theoduloz & Parra 1984).

Habitat: Chapito pool (October 2016 and May 2017), "small dam" (October 2016).

Closterium libellula Focke ex Nordstedt 1873

Cells right, fusiform 170-181.7 µm long, 40-41.7 µm wide. Chloroplasts with 5-12 longitudinal ridge, 3-6 pyrenoids. Cell wall smooth.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016), "small dam" (February 2017).

Closterium navicula (Brébisson) Lütkemüller 1905

Cells small, right, fusiform 74-80 µm long, 17-20 µm wide; apices broad, rounded to subtruncate. Terminal vacuoles spherical with 4 crystals. Chloroplasts 3-5 longitudinal ridge, 1-3 pyrenoids in each semicell. Cell wall smooth (Fig. 3f).

Recorded for the first time for Chile.

Habitat: Chapito pool (October 2016, February and August 2017).

Closterium parvulum Nägeli 1849

Cells strongly curved, 136 µm long, 15 µm wide. Chloroplasts with 2-3 longitudinal ridges, 2-5 pyrenoids in each semicell. Cell wall smooth.

Recorded for Chile (see Almanza *et al.* 2016; EULA 2016; Theoduloz & Parra 1984; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, February and August 2017), "small dam" (May 2017).

Closterium praelongum Brébisson 1856

Cells long, narrow, slightly curved 360-430 µm long, 20 µm wide; apices with inverse curvature. Chloroplasts with 3 longitudinal ridges and several pyrenoids. Cell wall finely striated.

Recorded for Chile (see Williamson 2004; Parra *et al.* 1983b).
Habitat: "small dam" (October 2016 and February 2017).

Closterium ralfsii Brébisson ex Ralfs 1848

Cells elongate-cylindrical, slightly curved, 413-511 µm long, 33-40 µm wide. Chloroplast 2-3 longitudinal ridges, 6-9 pyrenoids in each semicell. Cell wall striated, girdle bands present.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (February and August 2017).

Closterium striolatum Ehrenberg ex Ralfs 1848

Cell lightly curved, 276 µm long, 32 µm wide. Chloroplasts 5-6 longitudinal ridges, 5-9 pyrenoids in each semicells. Cell wall markedly striated with transverse bands.

Recorded for Chile (see Williamson 2004; Parra *et al.* 1983b).

Habitat: "small dam" (October 2016).

Cosmarium binum Nordstedt 1880

Cells large, 64-71 µm long, 48-54 µm wide. Semicells hemispherical – trapezoid; lateral margins with 9 crenations, 5 across the apex; crenations bigranulate. The protuberance in the middle region of the semicell bears 7 vertical rows of granules, and between this protuberance and the isthmus a row of large granules. Sinus deep closed linear; isthmus 13 µm wide.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016 and August 2017).

Cosmarium aff. bipunctatum Børjesen 1891

Cells small, almost as broad as long, 25.4 µm long, 23.6 µm wide. Semicells widely trapezoid, lower and upper angles obtuse, sides convex, apex widely truncated, margin crenate with 6 crenation at the apex and 6 along each side. Sinus linear, narrow gently dilated at the end; isthmus 7.8 µm wide (Fig. 3g). This taxon closely resembles *Cosmarium bipunctatum* Børjesen; however, it was impossible to observe, in the analyzed specimens, the two granules transversely arranged in the center of each semicell.

Recorded for the first time for Chile.

Habitat: Chapito pool (August 2017).

Cosmarium connatum Brébisson ex Ralfs 1848

Cells large 64-66 µm long, 48-49 µm wide. Semicells elliptical, apex broadly rounded. Isthmus 45-46 µm wide. Cell wall covered with small pores interspersed with many dots.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016 and August 2017).

Cosmarium laeve Rabenhorst 1868

Cells small, 25 µm long, 19 µm wide. Semicells truncated-pyramidal, flat apex. Sinus deep, narrow, linear; isthmus 8 µm wide. Cell wall scarcely punctate.

Recorded for Chile (see EULA 2013; Theoduloz & Parra 1984; Parra *et al.* 1983b).

Habitat: Chapito pool (August 2017).

Cosmarium portianum Archer 1860

Cells 41.3 µm long, 23.9 µm wide. Semicells oblong-ellipsoidal, reniform. Sinus deep, open; isthmus elongated 13 µm wide. Cell wall with granules.

Recorded for Chile (see Williamson 2004; Zúñiga *et al.* 1986; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, February and August 2017).

Cosmarium pseudopyramidatum P. Lundell 1871

Cells 59.2-62.4 µm long and 38.8 µm wide. Semicell pyramidal or semi-elliptical with flattened apex, basal angles rounded-angular. Sinus deep, narrow, open internally; isthmus 11.5-12.9 µm wide. Cell wall punctate.

Recorded for Chile (see, Zúñiga *et al.* 1986; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, August 2017).

Cosmarium pseudoretusum F. Duceillier 1918

Cells of medium size, 29-31µm long, 22-25µm wide. Semicell pyramidal, truncated apex, lateral margins with a basal lobe projecting outward, irregularly 2-undulate. Sinus straight, narrow; isthmus 5-6 µm wide. Cell wall punctate (Fig. 3h).

Recorded for the first time for Chile.

Habitat: Chapito pool (October 2016, February and August 2017).

Cosmarium reniforme (Ralfs) Archer 1874

Cells 48.8-50 µm long, 30.5-32.17 µm wide. Semicells reniform. Sinus deep, narrow; isthmus 11.93 µm wide. Cell wall granulate.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, February and August 2017).

Cosmarium sportella Brébisson ex Kützing 1849

Cells slightly longer than broad, 55-60 µm long, 48.5-51 µm wide. Semicell shortly pyramidal-truncated, with lower angles broadly rounded, apexes widely truncate and straight. Sinus deep, narrow, linear; isthmus 16.1 µm wide. Cell wall granulate, about 8-9 granules visible on each lateral margin, apex smooth or with 5 small marginal granules (Fig. 3i).

Reported for the first time for Chile

Habitat: Chapito pool (October 2016).

Cosmarium subprotumidum Nordstedt 1876

Cells small 24-25.4 µm long, 17.3-18 µm wide. Semicells trapezoidal with flattened apex and crenate lateral sides. Cell wall granulate.

Reported for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (February 2017).

Cosmarium subtumidum Nordstedt 1878

Cells 35-36 µm long, 26-28 µm wide. Semicell semicircular-pyramidal with broadly rounded basal and apical angle; apex widely truncated. Isthmus 9.8 µm wide. Cell wall finely punctate, but looks smooth.

Reported for Chile (see Almanza *et al.*, 2016; Parra *et al.* 1983b).

Habitat: Chapito pool (February and August 2017).

Cosmarium aff. wenmaniae Croasdale 1968

Cells 53.1-56.6 µm long, 52.3-55.3 µm wide. Semicells reniform with cell margin faintly crenulate. Sinus open abruptly and widely broad; isthmus 24.3 -29.8 µm wide. Cell wall rather thick and evenly coarsely scrobiculate. Chloroplast with two pyrenoids per semicell (Fig. 3j).

Reported for the first time for Chile.

Habitat: Chapito pool (October 2016, August 2017).

Desmidium grevillei (Kützing ex Ralfs) De Bary 1858

Cells 17-25 µm long, 34-42 µm wide, with an oval contour in front and rectangular in side view, which join together forming twisted filaments.

Reported for Chile as *Desmidium cylindricum* Greville (see Parra *et al.* 1983b).

Habitat: "small dam" (May 2017).

Euastrum binale (Turpin) Ehrenberg ex Ralfs 1848

Cells small, 19.7-20.8 µm long, 15.5-16.5 µm wide. Sinus deep, closed, linear; isthmus 4-4.4 µm wide. Semicells trapezoid-pyramidal-truncated; apex with a shallow open V-shaped incision marked by acute angles. Cell wall smooth.

Reported for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (August 2017).

Euastrum oblongum Ralfs 1848

Cells oblong-elliptical, 150-172.7 µm long, 70-92 µm wide. Sinus deep, narrow, linear; isthmus 21.8-30.9 µm wide. Semicells 5-lobed (one terminal and four lateral). Cell wall punctate.

Reported for Chile (see Parra *et al.* 1983b).

Habitat: "small dam" (October 2016, February 2017).

Gonatozygon aculeatum W.N. Hastings 1892

Cells cylindrical, 160-250 µm long, 14-20 µm wide. Cells unite loosely with each other to form filaments. Cell wall covered with fine spines 5-6.15 µm long.

Reported for Chile (see Pereira *et al.* 2000; Zúñiga *et al.* 1986; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, February, May and August 2017), "small dam" (February 2017).

Gonatozygon pilosum Wolle 1882

Cells cylindrical, straight, 81.5-103.4 µm long, 6.7-13.3 µm wide. Wall densely covered with small straight spines.

Reported for Chile (see EULA, 2011; Parra *et al.* 1983b, 1999, 2005).

Habitat: Chapito pool (October 2016, February and August 2017), "small dam" (May 2017).

Hyalotheca dissiliens Brébisson ex Ralfs 1848

Cells short slightly contracted in the medium 11-17 µm long, 21-23 µm wide, joined by truncated apices into long filaments, with mucilaginous sheath.

Reported for Chile (see Pereira *et al.* 2000; Theoduloz & Parra 1984; Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016, August 2017).

Micrasterias americana Ehrenberg ex Ralfs 1848

Cells compressed dorsoventrally, 118.9-127.4 µm long, 102.7 µm wide, deeply constricted with an open sinus. Semicells 5-lobed, polar lobe with two smaller ones that are arranged alternating one forward and one backward, separated from the lateral lobes by a deep incision; lateral lobes divided into two lobules with shallow incisions dissecting them. Cell wall granulate (Fig. 3k).

Recorded for the first time for Chile.

Habitat: Chapito pool (October 2016, February, May and August 2017).

Netrium interruptum (Brébisson ex Ralfs) Lütkemüller 1902

Cells cylindrical, 190 µm long, 36.6 µm wide, abruptly narrow towards the apexes. Vacuoles globular, large, terminal, with 1 crystal each. Chloroplasts are subdivided transversely giving the impression of 4 per cell.

Recorded for Chile (see Parra *et al.* 1983b)

Habitat: Chapito pool (February 2017).

Netrium oblongum (De Bary) Lütkemüller 1902

Cells oblong-elliptical with widely rounded ends, 150 µm long, 46 µm wide. Chloroplast lobed and striated with longitudinal and moderately dissected ridges, 1-2 pyrenoids.

Recorded for Chile (see Parra *et al.* 1983b)

Habitat: Chapito pool (October 2016).

Pleurotaenium ehrenbergii (Ralfs) De Bary 1858

Cells straight, cylindrical, 270-290 µm long, 30-35 µm wide. Semicells with a small basal inflation, apex truncate with 4 to 5 rounded tubercles. Chloroplasts narrow longitudinal bands, each with a row of small pyrenoids.

Recorded for Chile (see Williamson 2004; Pereira *et al.* 2000; Parra *et al.* 1983b).

Habitat: Chapito pool (February 2017), Tepual stream (August 2017).

Staurodesmus dejectus (Brébisson) Teiling 1954

Cells tri-radiate, 23.6-26.3 µm long, 20.9-25.4 µm wide. Semicells cup-shaped, apices with a short spine projected upwards. Isthmus 5-10 µm wide.

Recorded for Chile (see EULA 2011, 2012; Parra *et al.* 1983b, 1999, 2003, 2005; Theoduloz & Parra 1984).

Habitat: Chapito pool (October 2016, February and May 2017).

Staurodesmus dickiei (Ralfs) S. Lillieroth 1950

Cells tri-radiate, 29 µm long, 23.6-27.2 µm wide. Semicells elliptical with short, convergent spines. Isthmus 7.2-8.1 µm wide.

Recorded for Chile (see EULA 2012; Parra *et al.* 1983b, 1999, 2005; Zúñiga *et al.* 1986).

Habitat: Chapito pool (October 2016 and August 2017).

Staurodesmus glaber (Ralfs) Teiling 1948

Cells 2-3-4 radiate, 21.8-36.6 µm long, 15.4-35.4 µm wide. Semicells inverse triangles, angles with short incurved spines. Isthmus 6.3-9 µm wide.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016 and February 2017).

Staurastrum brachiatum Ralfs ex Ralfs 1848

Cells 27-36.5 µm long, 25-48 µm wide. Sinus acute, almost rectangular; isthmus 9 µm wide. Semicells triangular, arms or processes of variable length, with bifurcated ends. Cell wall smooth. Polar view 3 radiated.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (October 2016).

Staurastrum dilatatum Ehrenberg ex Ralfs 1848

Cells without arms or processes, 29-30 µm long, 25 µm wide. Semicells elliptical, upper margin slightly convex, lower inflated, lateral angles rounded or truncate-rounded. Isthmus 6.8-10 µm wide. Cell wall with small granules. Polar view 4-radiate.

Recorded for Chile (see Parra *et al.* 1983b).

Habitat: Chapito pool (May 2017).

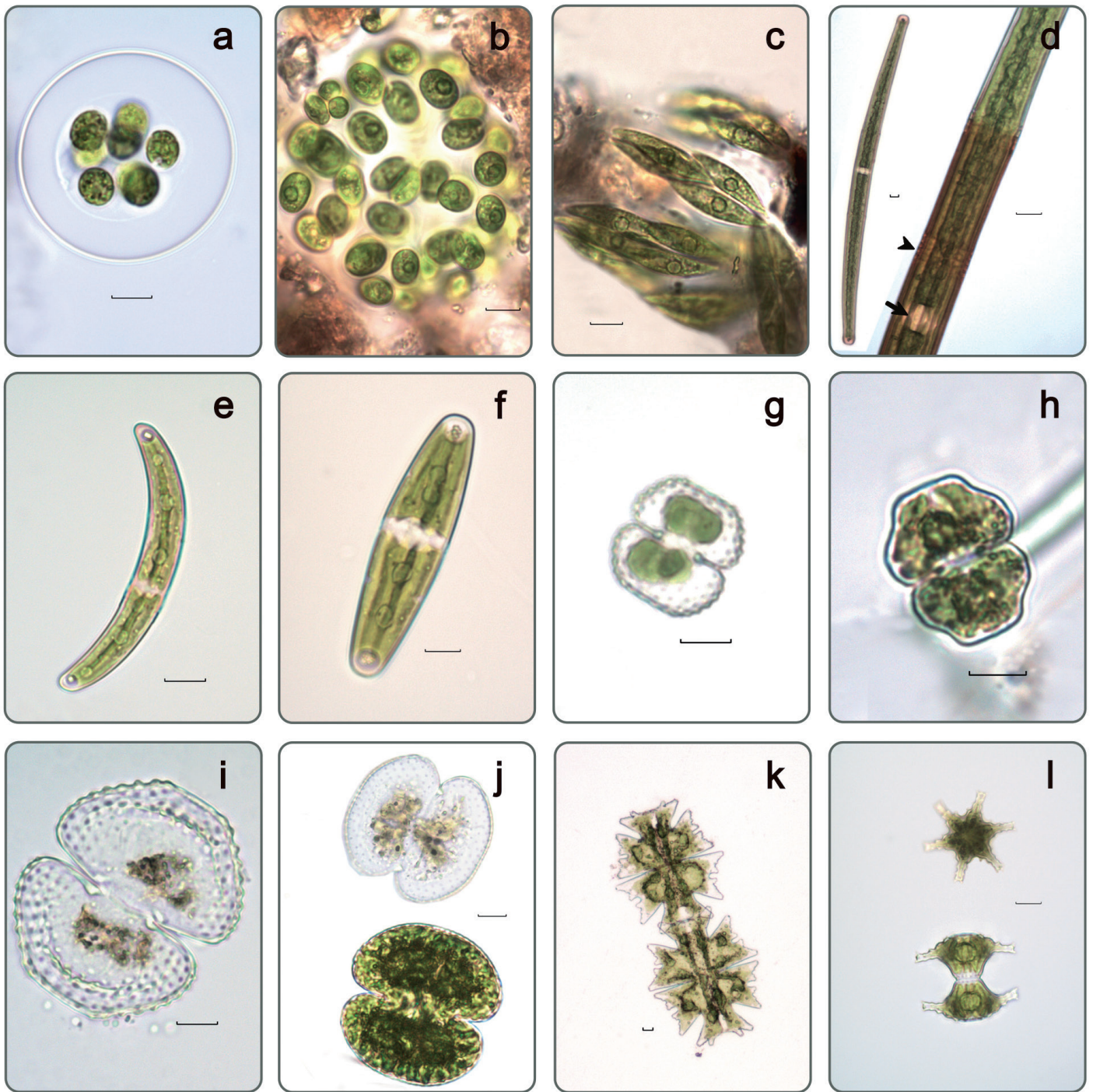


FIGURE 3. (a) *Chlamydocapsa* cf. *lobata*; (b) *Chlamydocapsa* cf. *planctonica*; (c) *Fusola* cf. *viridis*; (d) *Closterium angustatum*, ribs (arrow), bands (arrowhead); (e) *Closterium jeneri*; (f) *Closterium navicula*; (g) *Cosmarium* aff. *bipunctatum*; (h) *Cosmarium pseudoretusum*; (i) *Cosmarium sportella*; (j) *Cosmarium* aff. *wenmaniae*; (k) *Micrasteria americana*; (l) *Staurastrum margaritaceum*. Scale bar: 10 μ m. / (a) *Chlamydocapsa* cf. *lobata*; (b) *Chlamydocapsa* cf. *planctonica*; (c) *Fusola* cf. *viridis*; (d) *Closterium angustatum*, costas (flecha), bandas (punta de flecha); (e) *Closterium jeneri*; (f) *Closterium navicula*; (g) *Cosmarium* aff. *bipunctatum*; (h) *Cosmarium pseudoretusum*; (i) *Cosmarium sportella*; (j) *Cosmarium* aff. *wenmaniae*; (k) *Micrasteria americana*; (l) *Staurastrum margaritaceum*. Barra escala: 10 μ m.

Staurastrum furcigerum (Brébisson) W. Archer 1861

Cells 40-55.2 µm long, 47.8-55 µm wide (with processes). Semicells elliptical with short processes with concentric series of teeth, and a pair of extra-large teeth at the end. Isthmus 14.7-15 µm wide. Polar view 3-radiate.

Recorded for Chile (see Parra *et al.* 1983b, 1999, 2005; Theoduloz & Parra 1984).

Habitat: Chapito pool (August 2017).

Staurastrum manfeldtii Delponte 1877

Cells, 38 µm long, 40 µm wide (with processes). Semicells cup-shaped, processes horizontal or slightly convergent ending in 3 spines. Isthmus 10 µm wide. Cell wall smooth, but with granules on each process. Polar view 3-radiate.

Recorded for Chile (see Zúñiga *et al.* 1986; Parra *et al.* 1983b).

Habitat: Chapito pool (February 2017).

Staurastrum margaritaceum Meneghini ex Ralfs 1848

Cells 32-39 µm long and 36-44 µm wide (with processes). Semicells cup-shaped, with short processes that end in 4 small spines. Isthmus 10 µm wide. Cell wall with granules, with a concentric ring of granules in the isthmus. Polar view 6-radiate (Fig 3l).

Recorded for Chile (see Williamson 2004).

Habitat: Chapito pool (October 2016, February, May and August 2017).

Staurastrum rotula Nordstedt 1869

Cells 46.1-50 µm long, 26.1-27.7 µm wide; 73.9 µm wide (with processes). Isthmus 14.6 µm wide. Semicells sub-hexagonal; processes relatively long covered with small spines and ending in three teeth. Polar view 9-radiate.

Recorded for Chile (see González & Inostroza 2017; EULA, 2012, 2016).

Habitat: Chapito pool (May 2017).

Staurastrum sebaldi Reinsch 1866

Cells, 50-54 µm long and 58-75 µm wide (with processes). Isthmus 13-15 µm wide. Semicells cup-shaped; processes convergent, short, with warts and spines, ends truncated with 3-5 spines. Polar view 3-radiate.

Recorded for Chile (see Almanza *et al.* 2016; Asprey *et al.* 1964).

Habitat: Chapito pool (February and August 2017).

Staurastrum subavicularia (West) West & G.S. West 1894

Cells without arms, 31.8 µm long and 33.6 µm wide (with spines). Isthmus 12 µm wide. Semicells sub-elliptical; apices with a circle of spines and margins with 5-6 bifid spines. Polar view triangular.

Recorded for Chile (see Parra & González 1977).

Habitat: Chapito pool (October 2016, May and August 2017).

Xanthidium antilopaeum Kützing 1849

Cells 67 µm long (with spines), 48 µm (without spines). Semicells hexagonal, each of the four lateral angles extended into a pair of two simple spines. Cell wall finely punctate.

Recorded for Chile (see Parra *et al.* 1983b, 1999, 2003, 2005, Williamson 2004).

Habitat: "small dam" (February 2017).

The filamentous Zygnematalean genera *Mougeotia*, *Spirogyra* and *Zygnema* were also present in Chapito pool most of the year, but because no reproductive structures (zygotes) were found, it was not possible to identify them at species level, so were not included in this study. Most of the taxa identified were present in the Chapito pool and the "small dam". The Tepual stream was inhabited mainly by diatoms (not included in this study) and occasionally by some filamentous Zygnematalean (i.e. *Zygnema* sp.). No other group of microalgae was found in this lotic environment.

DISCUSSION

Most of the species were found inhabiting the Chapito pool (63 species- 77.7%); 8 species (9.8%) only in the "small dam" and ten species (12.3%) were present in both water bodies. On the other hand, few species (mostly diatoms, not included in this study) were present in the Tepual stream, probably because it was a lotic environment with fast moving water.

The range of pH values found during the annual recollection period (5.0 to 6.1) denote the presence of acidophilus water (pH <6.5, Šťastný 2010), a suitable environment for desmids which were the main group present in the water bodies studied. This group of greens occur exclusively in freshwater habitats, with a clear preference for oligo- to mesotrophic, slightly acidic environments (Kouwets 2008). According to this author, the largest species diversity can be found in shallow, mesotrophic lakes with rich stands of submerged macrophytes such as *Myriophyllum*, *Potamogeton* and *Utricularia*.

The 70.2 % of the desmids species identified in this investigation are from acidophilus environments (Šťastný 2010); on the other hand, the 48.6 % are indicators of mesotrophic environments and 21.6% of meso-oligotrophic habitats (Šťastný 2010). The most common species in term of presence and relative abundance was *Closterium angustatum*, a taxon characteristic for acidic and oligo-mesotrophic environments according to Šťastný (2010). They followed

it, in term of presence, *Staurastrum margaritaceum* (acid-oligo), *Micrasterias americana* (acid-meso) and *Gonatozygon aculeatum* (acid-meso). Within the other taxonomic groups, the blue-green *Chroococcus minutus* is known to inhabit mostly in shallower oligotrophic water bodies (Whitton 2008), the euglenoid *Euglena mutabilis* prevails in acidic habitats (Wolowski 2008) as well as the yellow-green flagellate *Vacuolaria virescens* (Pentecost 2008).

Even though all the samples come from phytoplankton, only few species of desmids are truly planktonic (Brook 1959). Brook stated that most species of genera *Closterium* and *Cosmarium* are confined to benthic habitat, while those belonging to *Staurastrum* are among the more numerous of planktonic desmids. Thus, many of the desmids encounter within this study are tychoplanktonic (dislodge by chance from macrophytes and get into the net) or benthic (may get into the net when dragged it close to the bottom) in their habitat, taking into consideration that Chapito pool is a very shallow water body. The same is true for most of the Oscillatoriales species and *Pseudanabaena* found, which are known from benthic, tychoplanktonic or periphytic environments.

Taxonomically, desmid taxa are generally distinguished on the basis of a large variety of more or less clearly defined morphological characters, mainly relating to cell wall outline and sculpture (Kouwets 2008). However, the taxonomic value of these characters, especially when they show a different degree of development, depend on the hierarchical level to be used. Their use at infraspecific taxonomic level (i.e. subspecies, varieties, forms) could lead to inconsistencies according to Kouwets (2008). Most of the genera found in this investigation are monophyletic (i.e. *Closterium* -Denboh *et al.* 2001, *Staurastrum* - Gontcharov & Melkonian 2005, *Gonatozygon* - Gontcharov *et al.* 2004, *Micrasterias* - Škaloud *et al.* 2011, *Bambusina* - Gontcharov & Melkonian 2011, *Pleurotaenium* - Škaloud *et al.* 2012 and *Xanthidium* - Stastný *et al.* 2013), and the species found could be easily identified since their morphological characters are prominent, well defined and clearly visible under the light microscopy. On the other hand, in *Cosmarium*, a polyphyletic genus according to Gontcharov & Melkonian (2011, 2008), the identification was harder because some characters were inconspicuous and less visible (such as cell wall sculpturing and pore partner). The taxonomy of *Cosmarium* are not resolved yet, so that any of the 12 species identify here may belong to this genus or may be included into new genera.

Within the Chlorophyta Chlorophyceae, the genera *Asterococcus* and *Chlamydocapsa* were very common in Chapito pool. No molecular studies have been done in either genera. Based on the study of Nakazawa *et al.* (2004) on *Asterococcus*, two species were identified in this investigation:

A. cf. limneticus and *A. cf. superbus*, mainly based on cell organization (single or in groups) and presence or absence of striated sheath around the cells. For *Chlamydocapsa*, Ettl & Gärtner (1988) recognized six species. Pröschold (2018, pers. comm.) identified as *C. cf. lobata* one of the species found in this study. The other species probably corresponds to *C. planctonica* since morphology fits well with the description of the life cycle of this taxon described by Skuja (1948).

Fusola viridis was found only in one occasion in Chapito pool (May 2017), and is the type and the only species of the genus *Fusola* Snow 1903. *Elakatothrix viridis* (J.W.Snow) Printz (1914), was the known name assigned to this microalga, but currently is regarded as its synonym (Guiry & Guiry 2018). *F. viridis* is nowadays a species accepted taxonomically. No molecular studies exist on neither organism, so their real taxonomic positions are still uncertain.

Most of the euglenoids taxa were found in Chapito pool, which could indicate the presence of organic matter in that water body with the subsequent bacterial activity.

Within the cyanobacteria, the most difficult group in term of taxa identification based on morpho-cytological attributes were the filamentous, and among them, the genera *Planktothrix*, *Tychonema* and *Anagnostidinema*. Two species of *Planktothrix* were present in the water bodies studied: *P. cf. clathrata* and *P. cf. suspensa*. According to Komárek & Komárková (2004), the infrageneric classification within *Planktothrix* is complex. Thus, *P. clathrata* forms a group with *P. compressa*. Both species belong to the "thicker" types and are distinguished only slightly by dimensions, so they are probably conspecific (Komárek & Komárková 2004). The same occurs with *P. suspensa* which form a cluster with *P. agardhii* (Komárek & Komárková 2004) and according to Davis *et al.* (2003), the separation of these species is doubtful according to present criteria.

Tychonema bornetii was only found once, but abundantly, in Chapito pool (August 2017). Only the type species of the genus (*Tychonema tenue* (Skuja) Anagnostidis & Komárek 1988), based on molecular phylogeny (16S rDNA), supports taxonomically the genus, since other species studied (*T. bornetii* not included) reveal that the genus is poly/paraphyletic.

Anagnostidinema is a recently described new genus erected by Strunecký *et al.* (2017), in order to include species previously assigned to *Geitlerinema* that differ from its congeneric taxa in the absence of capitate apical cells, and their phylogenetic molecular placement according to the 16S rRNA gene sequences. The morphological attributes exhibited by the individuals observed in this research, even though identified at first as *Geitlerinema cf. unigranulatum*, [= synonymous of *Geitlerinema amphibium* (Bittencourt-Oliveira *et al.* 2009)] fits well with the genus *Anagnostidinema* (apical

cells cylindrically rounded) and with the species *A. amphibium*, but molecular studies are needed to validate its identity.

One species of the genus *Oscillatoria* (*O. curviceps*) was also present and easily identified by morphological attributes. However, according to Komárek *et al.* (2014) this genus requires a taxonomic revision since molecular studies of some species have shown that the genus is paraphyletic or polyphyletic. They recall a lack of molecular data for most species of the genus.

Among the coccoid cyanobacteria, the identification was not as difficult as the previous group. The genus *Microcystis* was present in Chapito pool with two species: *M. natans* and *M. firma*. The genus is monophyletic (Komárek *et al.* 2014, Komárek & Komárková 2002) and its traditional species even though showed negligible molecular differences, differ in their morphology (*i.e.* coenobial form, mucilage structure, cell diameter, density of cells within the coenobium) and ecophysiology, so according to those authors, the traditional morpho-species category should be maintained.

Even though, Komárek *et al.* (2014) have categorized the genus *Aphanocapsa* as those morpho-genera that need taxonomic revision, the two species found in this study (*A. incerta* and *A. elachista*) were clearly differentiated by morphological attributes.

The recently erected genus *Anathece* (Komárek *et al.* 2011, originally a sub genus of *Aphanothece*) was also found in Chapito pool (August 2017) represented by one species: *A. minutissima*. However, the taxonomic stability of this genus is not completely supported, since the same authors found a high genetic similarity (16S rRNA) between species assigned to *Anathece* (coenobial) and to *Cyanobium* (unicellular), and they argue that their morphological differences may be a physiological expression to herbivory and/or to high irradiance. The above authors agree that more studies are needed to decide if both genera can be fused into only one genus.

The two species of *Pseudanabaena* inhabiting the Chapito pool were *Ps. catenata* and *Ps. minima*. According to Komárek & Anagnostidis (2005), the genus is polyphyletic, and there is not a clear separation of this genus with *Limnothrix*. Both genera need to be studied using polyphasic approach (Komárek *et al.* 2014).

Within Chroococaceae, two genera inhabit these water bodies: *Chroococcus* (*Chr. minutus*) and *Limnococcus* (*L. limneticus*). Both genera are currently recognized as distinct genera (Guiry & Guiry 2018), and the species found here were easily identify by their morphological attributes (cells disposition within the coenobium and amplitude of the mucilaginous envelope around the cells).

The only dinoflagellate commonly present in the water

bodies (Chapito pool, pH ranged from 5.0 to 5.8) within the park was *Parvodinium* (probably *P. cf. umbonatum* (F. Stein) Carty according to Ascencio, pers.observ. 2018). Carty (2014) considered this species as acid tolerant since she found it everywhere. The species was also reported as dominant in acidic lakes (Znachor & Nedoma 2014; Pérez *et al.* 1994). *Parvodinium umbonatum* is considered a species complex, whose circumscription is in progress (Kretschmann *et al.* 2018, Moestrup & Calado 2018).

Also, *Vacuolaria virescens* was the only Raphidophyceae inhabiting within the park and specifically in the “small dam”. This species tends to occur most commonly in small and usually rather acidic waters (Pentecost 2008), as it was in this water body (pH 5.8-6).

To conclude, this study enriches the biological knowledge already known from the park (*i.e.* trees, ferns, mosses, lichens, shrubs, birds, rodents, amphibian, some insect group), and constitute the first basic approach done on the photosynthetic biota of its water bodies. Only the Crustacean have been previously studied in the Chapito pool (De los Rios–Escalante & Ruedas 2014). The great diversity of desmids found is comparable with what Parra & González (1977) observed in 2 (out of 9) acidophilus freshwater bodies (Natri and Tarahuín lakes) from Chiloe Island, even though those water bodies were much bigger and deeper than the study sites. More than half (61.7%) of the species of desmids found in this research were also present in the nine water bodies from Chiloe Island. On the contrary, a much less desmids biodiversity (25 species) was found by Theoduloz & Parra (1984) in three acidophilus freshwater bodies (Lynch, Ojo del Mar and temporal Tres Puentes lagoons) from the extreme south of Chile, and there was only 10.6% of species coincidence. Finally, the taxonomic results denote the great diversity of microalgae inhabiting these pristine and peculiar environments and will serve as a basis for further studies (a) in similar ecological areas within the country and/or (b) focused on the taxonomy and systematics of a particular species or species complexes.

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